

# Cellufine™ GH-25

**Cellufine GH-25 provides a rapid means of salt removal and buffer exchange for protein solutions. The semi-rigid spherical cellulose beads allow high flow rates with little compression of the column bed.**

The separation mechanism is based on differential solute access into the chromatographic bead. Whereas large molecules (above 3 kD) are excluded from the packing and pass quickly through the column, small molecules (salts) diffuse into the bead and are thus retained longer. GH-25 can be used to remove alcohols, salts, detergents, fluorochromes, sugar, etc., from virtually any protein solution. It is compatible with most solvents and is stable from pH 1 - 14.

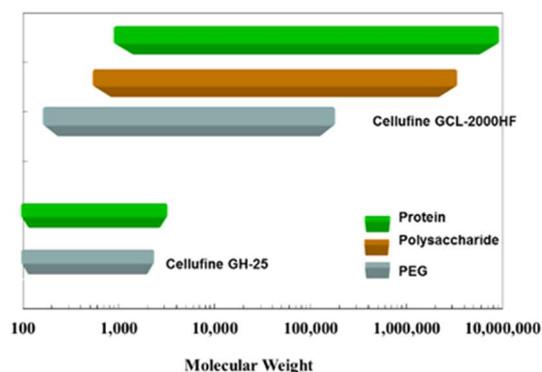


Figure 1 Fractionation range of Cellufine GH-25

Table 1. Characteristics of Cellufine GH-25

|                           |                              |
|---------------------------|------------------------------|
| <b>Matrix</b>             | Spherical cellulose particle |
| <b>Particle size</b>      | 40 - 130 μm (ca.90mm)        |
| <b>MW exclusion limit</b> | 3kD                          |
| <b>Operating pressure</b> | <0.2 MPa                     |
| <b>pH stability range</b> | 1 - 14                       |
| <b>Storage</b>            | 2-8 °C in 20 % ethanol       |

Values in Table 1 are not specifications.

## Column Packing

### Materials

- Cellufine GH-25
- Lab scale column, adapter, reservoir
- Pump
- Filtration equipment (Glass filter, Buchner-Roth, aspirator, etc.)
- Graduated cylinder
- Packing solution (water, NaCl solution※, buffer※)
- Mobile phase of packed column evaluation ( water, NaCl solution※, buffer※)
- Sample of packed column evaluation ( 1-2 %(v/v)acetone or 1M NaCl )

※NaCl solution : low salt concentration solution such as 0.1M NaCl solution

※buffer : Adsorption buffer , etc.

### Preparation of slurry

- 1) After the bottle is warmed to room temperature, shake it several times to equalize the slurry in the bottle.
- 2) Aspirate through a glass filter and wash 3 times with 5 times the volume of the packing solution. Remove 20% ethanol as a preservative. If necessary, the slurry may be washed by decantation.
- 3) Transfer the resin to a beaker and add packing solution to make a 50-60% (v/v)slurry. Suspend the resin and remove the air under decompression for 30-40 minutes. Slow stirring with a magnetic stirrer can effectively remove the air.

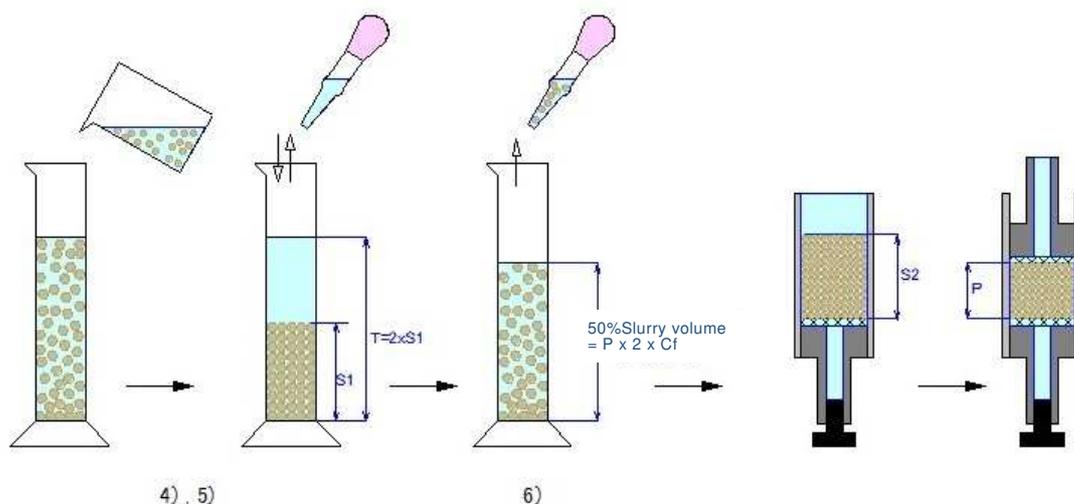


Figure2 Preparation of slurry

- 4) Pour the slurry into a graduated cylinder and leave it to stand for at least 4 hours. Measure bed height (volume) of a gravity settled bed and calculate the slurry concentration.

**Slurry concentration (%) =**

$$\text{Gravity settled bed volume (S1)} / \text{Total slurry volume (T)} \times 100$$

- 5) Adjust to a 50 % (v/v) slurry concentration of resin with packing solution.
- 6) Calculate the volume of slurry required to pack the column.

$$\text{50\% slurry volume required to packing} = (\text{Target packing volume (P)} \times 2) \times \text{Cf}$$

$$\text{※Cf} = [\text{gravity settled bed volume (S2)} / \text{Target packing volume (P)}]$$

**Note:** *Compression factor (Cf) affects the column packing efficiency (see Appendix 1 for details) and may need to be optimized by adjusting the axial compression on the column.*

### Column packing

- 1) Set up the column. After opening the column outlet, add packing solution to remove air remaining in the filter. Leave about 1 cm of packing solution from the bottom of the column to prevent air from entering.
- 2) Close the column outlet and pour the slurry into the column all at once to keep air out.
- 3) Open the column outlet to allow the resin to settle. As the resin settles, the liquid surface becomes clear. Close the outlet when the packing solution becomes clear to 2-3 cm from the liquid surface.
- 4) Carefully fill the column to the top with packing solution. Keep the settling resin from floating away.
- 5) Set the adapter on the column to prevent air from entering the liquid surface. Close the adapter O-ring, lower the adapter, and remove the air inside.
- 6) Connect the column to the pump and keep the packing solution flowing at a pressure lower than the operating pressure for 30 to 60 minutes. (<0.2MPa)

**Note:** *The flow velocity : Internal pressure at packing > Operating pressure after packing*

- 7) After the resin height is stabilized, stop the flow. Close the column outlet. Disconnect the inlet piping at the top of the column. Slowly lower the adapter to the surface of the resin. At this time, the packing solution in the column flows out from the column inlet.
- 8) Connect to the adapter inlet with the piping filled with packing solution to exclude air. Open the column outlet and pump the packing solution. (< 0.2MPa) If the resin compresses and makes space between it and the adapter, lower the adapter to the surface of the resin.
- 9) Calculate column volume from column height. If the column volume is larger than the target column volume, lower the adapter to the target height. If the column volume is lower than the target column volume, the slurry concentration is low, or the resin may be excessively compressed. Remove the resin from the column and pack again.

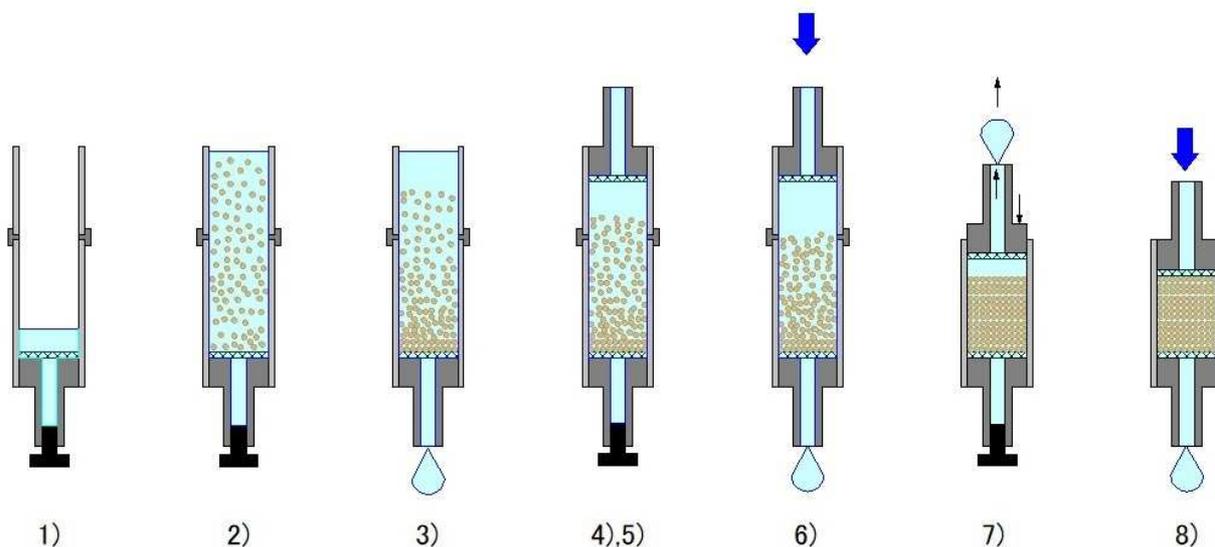


Figure 3 Process of column packing

## Evaluation of column packing

Column packing efficiency is evaluated by checking HETP and asymmetry (As).  
(Appendix 1)

## Operating Guidelines

### How to use

- 1) Equilibrate the column with equalization buffer. (3-5 column volume (CV))
- 2) Load the column with sample dissolved in equalization buffer.

- 3) Wash with equalization buffer to remove impurities. (2-3 CV)
- 4) Elute the adsorbed target substance with equalization buffer.

### Sample preparation and loading

Samples are typically loaded in the buffer which is to be exchanged. Filtration may be required to remove insoluble matter. The sample load is calculated as a function of column volume. Sample loads of 10% to 30% of total column volume are recommended. At higher loads, samples become less diluted. However, salt removal may not be absolute. Furthermore, volume loadability is inversely related to protein concentration.

### Recommended flow rate

100–300 cm/h. (<0.2MPa)

### Elution

Elution occurs under isocratic conditions. The protein and salt/alcohol should elute at approximately 30% and 85% of the total column volume, respectively.

### Stability

Stable in:

pH 1–14

Ethanol, methanol, acetone, etc.

8 M Urea, 6 M Guanidine/HCl

0.1 M HCl

0.5 M NaOH

Most salts (NaCl, (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, etc.)

Most detergents (SDS, Tween®, Chaps, etc.)

Autoclavable: 121°C at 1 bar for 20 minutes

### Regeneration

Flush the column with 2-5 bed volumes of 0.1 - 0.5 M NaOH at a velocity of 50 –100 cm/h. Remove caustic by flushing with several bed volumes of DIW or exchange buffer. In the

later case, measure the pH of the column eluate to ensure that the system has returned to equilibrium.

## Storage

Unopened resin should be stored at 2-8°C. After opened or packed columns are equilibrated with 20% ethanol and stored at 2-8°C. Do not freeze. 5 years from date of manufacture.

## Product Ordering Information

| Description        | Pack Size            | Catalogue No. |
|--------------------|----------------------|---------------|
| Cellufine<br>GH-25 | 5 mL×5 (Mini-column) | 19711-55      |
|                    | 100 mL               | 670 000 327   |
|                    | 500 mL               | 19711         |
|                    | 5 L                  | 19712         |
|                    | 10 L                 | 670 000 335   |

## Purchase/Technical Support

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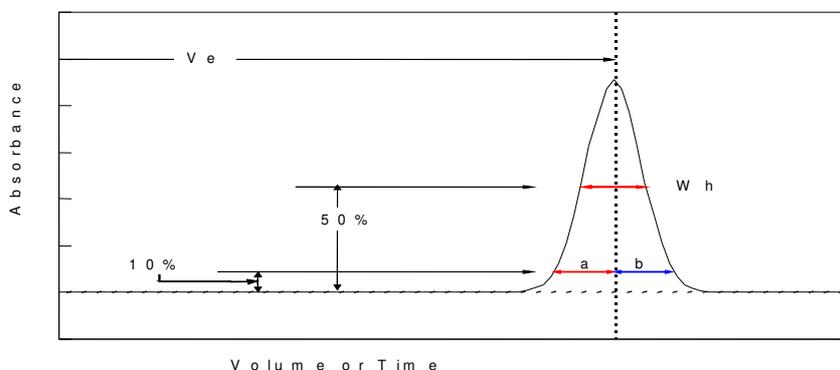
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### Appendix 1: Evaluation of column packing

The packing condition of the column is evaluated using indices such as theoretical number of stage plates (N), theoretical stage equivalent height (HETP), and asymmetry (As). These indices are affected by the measurement conditions. For example, these indices vary with column diameter/height differences, piping, solvent sample volume, flow rate, temperature, etc. Therefore, it is necessary to use the same measurement conditions each time. A flow velocity of 30 cm/h is recommended, but higher velocities are possible. However, the higher the flow velocity, the smaller the theoretical number of plates (N). The same conditions (flow rate, column size, mobile phase, and sample) must be used each time to evaluate the column.

| Parameter            | Condition  |
|----------------------|--|
| Sample volume        | 1 -2.5% of column volume (CV)                                  |
| Sample concentration | 1-2 %(v/v) acetone (mobile phase : water or adsorption buffer) |
|                      | 1M NaCl (mobile phase : 0.1-0.4M NaCl aq)                      |
| Flow rate (cm/h)     | 30 cm/h  |
| Detector             | UV, Conductivity   |

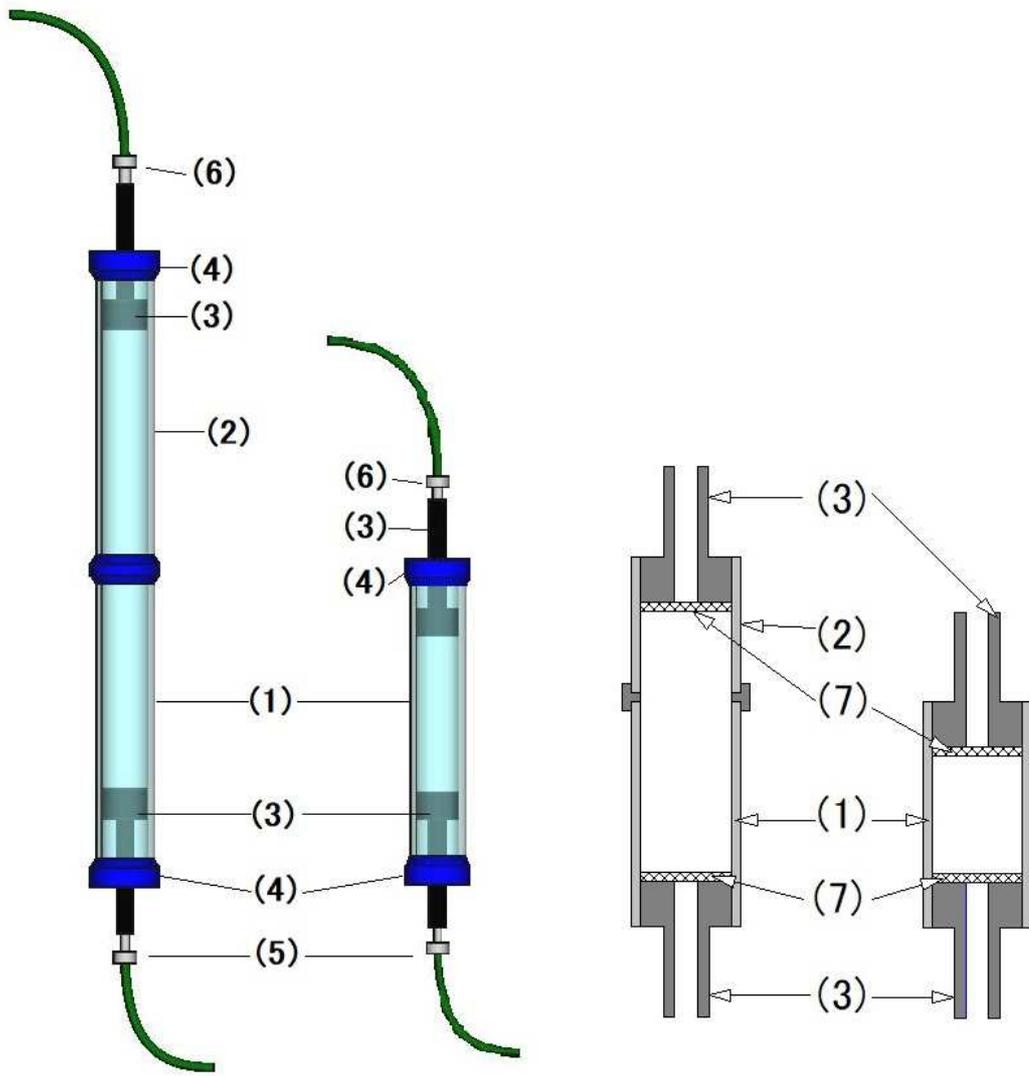


|       |  |
|-------|--|
| L     | Column length [cm or m]                                |
| $V_e$ | Elution time or volume                                 |
| $W_h$ | Half of width of peak                                  |
| a, b  | Peak width of 10% peak height<br>(a) front<br>(b) rear |
| Note  | $V_e, W_h$ and a, b should have same dimensional units |

$$\text{HETP} = L/N \quad N = 5.54 \times (V_e/W_h)^2 \quad A_s = b/a$$

Generally, number of (theoretical) plates (N) is good if it is over 3000. Acceptable asymmetry factor (As) values range from 0.7-1.5.

**Appendix 2 : Figure of columns**



The Operating Instructions use a simple cross-sectional figure of the column, as shown on the right.

|     |             |     |               |
|-----|-------------|-----|---------------|
| (1) | Column tube | (4) | Column end    |
| (2) | Reservoir   | (5) | Column outlet |
| (3) | Adapter     | (6) | Column inlet  |
| (7) | Filter      |     |               |