

## Operating Instructions

# Cation Exchange Chromatography Media Cellufine C-500

#### **Description**

Cellufine C-500 media manufactured by JNC Corporation are designed for the cation exchange chromatography of basic proteins and other biomolecules. The resins are comprised of beaded spherical cellulose, functionalized with carboxymethyl (CM). Cellufine C-500 medium is ideal for the chromatography of proteins molecular weight up to 500 kD. The superior rigidity of Cellufine gels allows for high flow rates, and thus rapid processing times, even in large diameter process scale columns.

# **Physical-Chemical Characteristics**

Support matrix	Cellulose		
Particle shape	Spherical		
Particle diameter (µm)	ca. 40 $-$ 130 (Ave.; 90 $\mu$ m)		
Ion capacity (meq/g dry)	0.9-1.2		
Swelling degree (ml / g dry media)	9–11		
MW exclusion limit (kD)	500		
pH stability range	1 – 13		
Operating pressure	< 2 bar (29 psi)		
Tapped volume (ml / g succession media)	1.3 – 1.5		
Supplied	suspension in 20 % EtOH		

#### Column Packing

- 1. Prepare a 40 60 % (v/v) slurry with water, 0.1 M NaCl solution or the appropriate buffer and allow to equilibrate at ambient temperature for one hour.
- 2. Gently stir or place under vacuum to degas.
- 3. With column outlet closed, carefully pour the slurry into the column. If necessary, fit column with a filler tube to accommodate the entire slurry volume.
- 4. Attach upper end cell to column, then pump 10 column volumes of elution buffer at a flow rate 20 % 50 % greater than the operational flow rate.
- 5. After flushing, remove filter tube and reattach end cell to the column tube.
- 6. Equilibrate with 10 15 column volumes of adsorption buffer in preparation for sample loading.



## **Operating Guidelines**

#### **General Operation**

Typically, adsorption to Cellufine Cation Exchange medium occurs in relatively low ionic strength (e.g., below 0.1 M NaCl) in the pH range from 5.0 - 7.0. Under these conditions, proteins with neutral or net positive charge will bind. Bound components are then resolved by stepwise elution with buffers containing progressively higher salt concentration or by elution with a linear salt gradient.

## Sample Preparation and Load

Prepare sample by centrifugation or microfiltration to remove insoluble material. Samples should be prepared to a concentration of 1 - 20 mg/ml, in binding buffer or at comparable conditions of ionic strength and pH. If necessary, exchange sample buffer using dialysis, diafiltration or desalting chromatography.

#### **Recommended Buffers**

Adsorption buffer: 0.02 – 0.05 M sodium acetate (pH 5.5)

Elution buffer: 0.1 - 2.0 M sodium chloride in adsorption buffer.

Other common buffer systems may be used. For additional information on chromatographic methods of protein purification, see References.

# **Chemical and Physical Stability**

Stable in:

Most salts (NaCl, (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, etc.) Most detergents (SDS, Tween®, Chaps, etc.) < 0.5 M NaOH

Autoclavable: 121°C at 1 bar (14.5 psi) for 20 minutes

Flow Rate

The recommended linear velocity range for Cellufine C-500 is 50 – 200 cm/h.

# Regeneration and Depyrogenation

To regenerate a column, flush bed with 2 - 5 column volumes of 0.5 N NaOH, followed by several volumes of elution buffer. Then equilibrate as usual. If the column needs to be pyrogen free, wash the column with 2 to 5 column volumes of 0.5 N NaOH followed by several column volumes of pyrogen free elution buffer. Monitor the pyrogen levels in the column eluate during a blank gradient elution prior to reusing the column.



#### **Storage**

Store unopened container at ambient temperature. Do not freeze.

Short term (2 weeks or less), bulk and column can be stored at a room temperature with 0.05 N NaOH. Longer storage should be in neutral buffer containing 0.02 % sodium azide or 20 % ethanol, at 2-8 °C. Do not freeze.

#### Shelf Lifetime:

5 years from date of manufacture

#### References

- 1. Harris, E.L.V. and Angal, S., *Protein Purification Methods: A practical Approach*. New York: Oxford University Press, 1989.
- 2. Janson, J.-C. and Ryden, L., *Protein Purification: Principles, High Resolution Methods, and Applications.* 2<sup>nd</sup> ed. New York: John Wiley & Sons, Inc., 1998

#### **Product Ordering Information** (Catalogue No.)

Media type	Pack Size					
	Mini-Column 1ml x 5	100ml	500ml	5 lt	10 lt	
Cellufine C-500	19800-51	675 983 327	19865	19866	675 983 335	
Cellufine Q-500	19907-51	675 982 327	19907	19908	675 982 335	
Cellufine A-200		676 980 327	19611	19612	676 980 335	
Cellufine A-500	19805-51	675 980 327	19805	19806	675 980 335	
Cellufine A-800		673 980 327	19800	19801	673 980 335	

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