

CellufineTM Phosphate

Cellufine Phosphate an affinity chromatography media designed for the concentration, purification protein kinase, nucleic acid related proteins such as restriction enzymes, nuclease, polymerase. This media is based on spherical cellulose beads functionalized with a phosphate ester.

Compared with the conventional product, Cellufine Phosphate has resistance to pressure and use in a large-sized column with high flow rate is also possible. This is because Cellufine Phosphate is a spherical particle, but conventional product is a fiber. Table 1 shows the characteristics of Cellufine Phosphate.

Table 1, Characteristics of Cellufine Phosphate

	Characteristics
Ligand	Phosphate ester
Matrix	Spherical cellulose particle
Particle size	40 - 130 μ m (ca.90 μ m)
MW exclusion limit	100 kD (PEG)
Ion exchange capacity (meq/mL-gel)	0.3 - 0.8
Operating pressure	<0.2 MPa
Protein Binding Capacity	> 20 mg /ml (Lysozyme)
pH stability range	5 – 12
Storage	2-8 °C in 20 % ethanol

※Values in Table 1 are not specifications.

Column Packing

Materials

- Cellufine resin
- Lab scale column, adapter, reservoir
- Pump
- Filtration equipment (Glass filter, Buchner-Roth, aspirator, etc.)
- Graduated cylinder
- Packing solution (water, NaCl solution※, buffer※)

- Mobile phase of packed column evaluation (water, NaCl solution※, buffer※)
- Sample of packed column evaluation (1-2 %(v/v)acetone or 1M NaCl)

※NaCl solution : low salt concentration solution such as 0.1M NaCl solution

※buffer : Adsorption buffer , etc.

Preparation of slurry

- 1) After the bottle is warmed to room temperature, shake it several times to equalize the slurry in the bottle.
- 2) Aspirate through a glass filter and wash 3 times with 5 times the volume of the packing solution. Remove 20% ethanol as a preservative. If necessary, the slurry may be washed by decantation.
- 3) Transfer the resin to a beaker and add packing solution to make a 50-60%(v/v) slurry. Suspend the resin and remove the air under decompression for 30-40 minutes. Slow stirring with a magnetic stirrer can effectively remove the air.

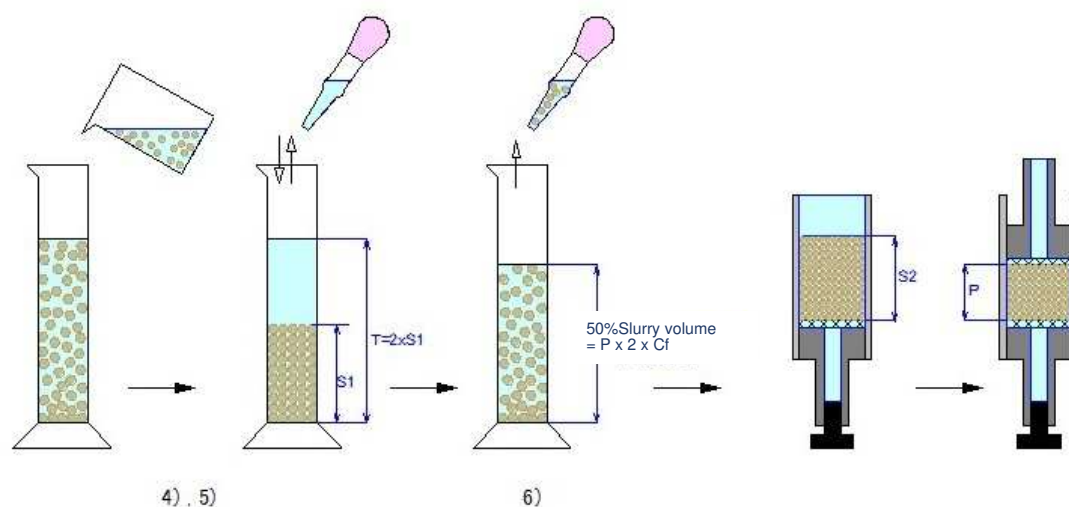


Figure1 Preparation of slurry

- 4) Pour the slurry into a graduated cylinder and leave it to stand for at least 4 hours. Measure bed height (volume) of a gravity settled bed and calculate the slurry concentration.

Slurry concentration (%)

$$= \text{Gravity settled bed volume (S1)} / \text{Total slurry volume (T)} \times 100$$

- 5) Adjust to a 50 % (v/v) slurry concentration of resin with packing solution.

6) Calculate the volume of slurry required to pack the column.

$$\text{50\% slurry volume required to packing} = (\text{Target packing volume (P)} \times 2) \times C_f$$

$$\times C_f = [\text{gravity settled bed volume (S2)} / \text{Target packing volume (P)}]$$

Note: Compression factor (C_f) affects the column packing efficiency (see Appendix 1 for details) and may need to be optimized by adjusting the axial compression on the column.

Column size (Diameter × Bed height)	Recommended C_f (approximately)
5.0 cm × 21.7 cm	1.30

Column packing

- 1) Set up the column. After opening the column outlet, add packing solution to remove air remaining in the filter. Leave about 1 cm of packing solution from the bottom of the column to prevent air from entering.
- 2) Close the column outlet and pour the slurry into the column all at once to keep air out.
- 3) Open the column outlet to allow the resin to settle. As the resin settles, the liquid surface becomes clear. Close the outlet when the packing solution becomes clear to 2-3 cm from the liquid surface.
- 4) Carefully fill the column to the top with packing solution. Keep the settling resin from floating away.
- 5) Set the adapter on the column to prevent air from entering the liquid surface. Close the adapter O-ring, lower the adapter, and remove the air inside.
- 6) Connect the column to the pump and keep the packing solution flowing at a pressure lower than the operating pressure for 30 to 60 minutes. (< 0.2MPa)

Note: The flow velocity : Internal pressure at packing > Operating pressure after packing

- 7) After the resin height is stabilized, stop the flow. Close the column outlet. Disconnect the inlet piping at the top of the column. Slowly lower the

adapter to the surface of the resin. At this time, the packing solution in the column flows out from the column inlet.

- 8) Connect to the adapter inlet with the piping filled with packing solution to exclude air. Open the column outlet and pump the packing solution. ($< 0.2\text{MPa}$)
- 9) If the resin compresses and makes space between it and the adapter, lower the adapter to the surface of the resin.
- 10) Calculate column volume from column height. If the column volume is larger than the target column volume, lower the adapter to the target height. If the column volume is lower than the target column volume, the slurry concentration is low, or the resin may be excessively compressed. Remove the resin from the column and pack again.

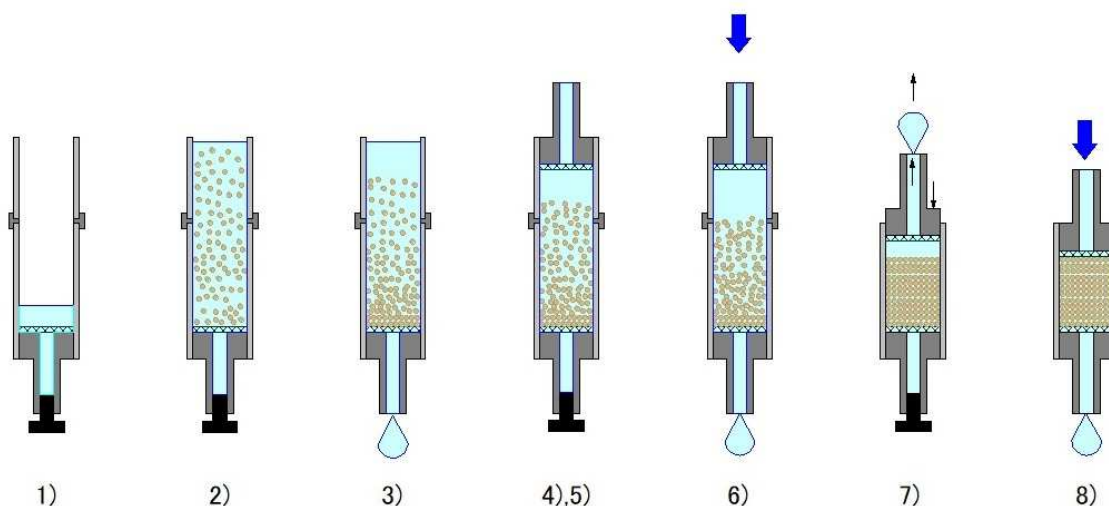


Figure 2 Process of column packing

Evaluation of column packing

Column packing efficiency is evaluated by checking HETP and asymmetry (As). (Appendix 1)

Operating Guidelines

How to use

- 1) Equilibrate the column with adsorption buffer. (3-5 column volume (CV))
- 2) Load the column with sample dissolved in adsorption buffer. (1-20mg/mL)
- 3) Wash with adsorption buffer to remove impurities. (2-3 CV)
- 4) Elute the adsorbed target substance with elution buffer.

Recommended Buffers

Adsorption buffer : 10mM to 50mM sodium phosphate, neutral pH. Depending on the application, other buffer ions such as tris, acetate, may be used. In general, adsorption strength varies inversely with pH and ionic strength. Increasing ionic strength slightly can aid in removing loosely bound contaminants. Non-ionic detergents (Tween®20, Triton® X, etc.) may also be added to improve solubility.

Elution buffer : In general use mobile phase consisting of adsorption buffer containing 1 – 2 M NaCl or KCl. The exact concentration can be determined by gradient elution. Step gradients are typically employed for preparative applications.

Sample Preparation and Load

Prepare samples at a concentration of 1 – 20 mg/ml, in adsorption buffer. Remove insoluble material by centrifugation or microfiltration. If necessary, exchange sample buffer using dialysis, diafiltration or desalting chromatography.

Flow Rate

The recommended linear velocity range for Cellufine Phosphate media is 50 – 200 cm/h.

Regeneration and Depyrogenation

Cellufine Phosphate is typically regenerated and depyrogenated with high ionic strength (2.0 – 3.0 M) NaCl. If this is not sufficient, regenerate more aggressively with 3 – 10 column volumes of 0.2 N NaOH at 2 – 10°C, then wash with 2.0 – 3.0 M NaCl until pH drops below 9. Wash gel again with starting buffer until equilibrated.

Stability

pH range of 5 to 12 and operating temperature of 2 to 30°C are recommended.

Storage

Store unused resin in its container at a temperature of 2 to 8°C. Equilibrate opened resin and packed column in 20% ethanol and store at 2 to 8°C. Can be

stored in buffer containing 1M NaCl in bulk and column conditions for up to 2 weeks. For long-term storage, add a preservative such as 0.02% sodium azide. Do not freeze. Shelf Lifetime is 5 years from manufacture.

References

Nucleic Acids Research, 2006, Vol. 00, No. 00 1–8

Rachel Macmaster, Svetlana Sedelnikova, Patrick J. Baker, Edward L. Bolt¹, Robert G. Lloyd¹ and John B. Rafferty

RusA Holliday junction resolvase: DNA complex structure—insights into selectivity and specificity

Product Ordering Information (Catalogue No.)

	Pack Size	Catalogue No.
Cellufine Phosphate	1 mL x 5 (Mini-Column)	19551
	5 mL x 1 (Mini-Column)	19515
	10 mL	19524
	50 mL	19545
	500 mL	19546
	5 L	684 987 330
	10 L	684 987 335

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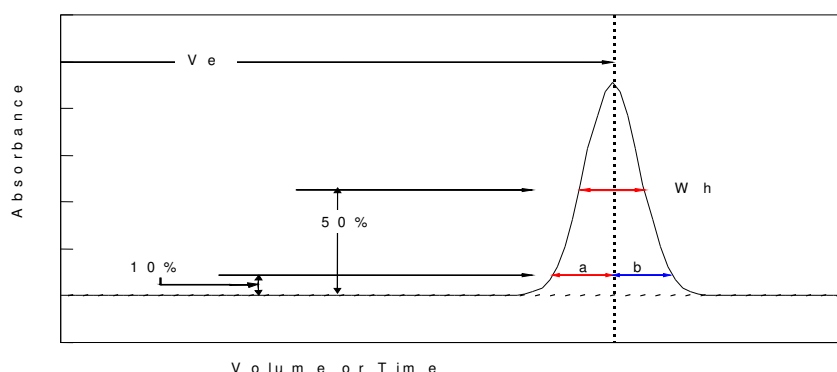
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Appendix 1: Evaluation of column packing

The packing condition of the column is evaluated using indices such as theoretical number of stage plates (N), theoretical stage equivalent height (HETP), and asymmetry (As). These indices are affected by the measurement conditions. For example, these indices vary with column diameter/height differences, piping, solvent sample volume, flow rate, temperature, etc. Therefore, it is necessary to use the same measurement conditions each time. A flow velocity of 30 cm/h is recommended, but higher velocities are possible. However, the higher the flow velocity, the smaller the theoretical number of plates (N). The same conditions (flow rate, column size, mobile phase, and sample) must be used each time to evaluate the column.

Parameter	Condition
Sample volume	1 -2.5% of column volume (CV)
Sample concentration	1-2 %(v/v) acetone (mobile phase : water or adsorption buffer)
	1M NaCl (mobile phase : 0.1~0.4M NaCl aq)
Flow rate (cm/h)	30 cm/h
Detector	UV, Conductivity



L	Column length [cm or m]
V_e	Elution time or volume
W_h	Half of width of peak
a, b	Peak width of 10% peak height (a) front (b) rear
Note	V_e, W_h and a, b should have same dimensional units

$$\text{HETP} = L/N$$

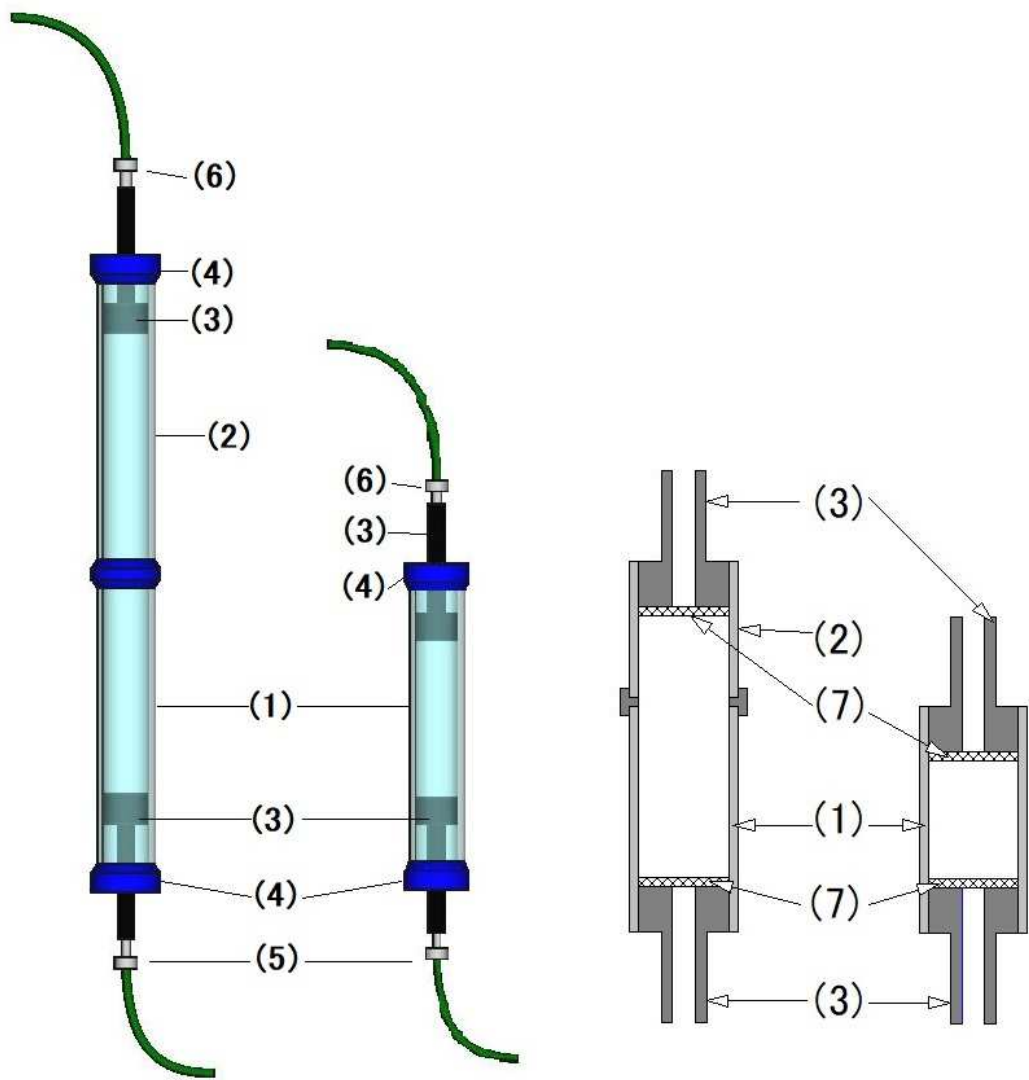
$$N = 5.54 \times (V_e/W_h)^2$$

$$A_s = b/a$$

Generally, number of (theoretical) plates (N) is good if it is over 3000.

Acceptable asymmetry factor (A_s) values range from 0.7-1.5.

Appendix 2 : Figure of columns



The Operating Instructions use a simple cross-sectional figure of the column, as shown on the right.

(1)	Column tube	(4)	Column end
(2)	Reservoir	(5)	Column outlet
(3)	Adapter	(6)	Column inlet
(7)	Filter		

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